Conformational Studies of (±)-11,12-Didehydro-11-deoxycorynoline and (+)-11,13-Didehydro-11-deoxychelidonine, Hexahydrobenzo[c]phenanthridine-Type Alkaloid Derivatives, by X-Ray Crystal Structure and NMR Analyses

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The X-ray crystal analyses of the two 11-deoxy-didehydrohexahydrobenzo[*c*]phenanthridine-type alkaloid derivatives **3** and **4**, derived from (\pm)-corynoline (**1**) and (+)-chelidonine (**2**), established their structures as (\pm)-(5b*RS*,12b*RS*)-5b,12b,13,14-tetrahydro-5b,13-dimethyl[1,3]benzodioxolo[5,6-*c*]-1,3-dioxolo[4,5-*i*]phenanthridine (**3**) and (+)-*rel*-(12b*R*)-7,12b,13,14-tetrahydro-13-methyl[1,3]benzodioxolo[5,6-*c*]-1,3-dioxolo[4,5-*i*]phenanthridine (**4**). The conformations of **3** and **4** in CDCl₃ were determined on the basis of ¹H- and ¹³C-NMR spectroscopy.

Introduction. – We have been investigating the relationship between structure and cell-adhesion-inhibitory activity of isoquinoline alkaloids [1][2]. Corynoline (1) [3] and chelidonine (2) [4], representative hexahydrobenzo[*c*]phenanthridine-type alkaloids [5], show a quite different activity for the cell-adhesion-inhibitory intensity [1], probably due to the presence or absence of the Me group at $C(13)^1$). A similar deviation has been observed for the corresponding dehydrated compounds, *i.e.*, (±)-11,12-didehydro-11-deoxycorynoline¹) (3) [6] and (+)-11,13-didehydro-11-deoxychelidonine¹) (4) [7][8], where **3** shows an inhibitory activity of *IC*₅₀ 166.0 µM which is slightly weaker than that of **1** (*IC*₅₀ 54.8 µM) [1][2], while **4** exhibits no inhibitory activity at all. These compounds were prepared by dehydration of the OH–C(11) groups of **1** and **2** (*Fig. 1*).

The cell-adhesion-inhibitory effect could be due to an interaction of the inhibitor with the adhesive molecule [9], such as ICAM-1 [10][11], on blood-vessel epithelial cells. Thus, it is important to elucidate the conformational feature of the inhibitor for elucidating the structure – activity relationship. The chemical structures of **3** and **4** were determined by spectral methods [6–8], but their conformational features are still unknown. To compare these conformations with those of **1** and **2** and to further accumulate informations on the structure – activity relationship of hydrobenzo[c]phenanthridine-type alkaloids, we analyzed the solid-state and solution conformations of **3** and **4**.

Results and Discussion. – The crystallographic data of **3** and **4** are shown in *Table 1*. Selected bond lengths, bond angles, and torsion angles can be found in *Table 2*.

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Fig. 1. Structures of 1-4, together with the atom numbering used in this work¹), and anti-cis and syn-cis conformations illustrated by Newman projections viewed from C(13) to C(14) of the B/C-cis-fused hexahydrobenzo[c]phenanthridine skeleton

The structure of **3** was determined by X-ray crystal analysis to be (\pm) -11,12didehydro-11-deoxy-13-methylchelidonine $(=(\pm)-(5bRS,12RS)-5b,12b,13,14$ -tetrahydro-5b,13-dimethyl[1,3]benzodioxolo[5,6-*c*]-1,3-dioxolo[4,5-*i*]phenanthridine), and that of **4** to be (+)-11,13-didehydro-11-deoxychelidonine (=(+)-rel-(12bR)-7,12b,13,14-tetrahydro-13-methyl[1,3]benzodioxolo[5,6-c]-1,3-dioxolo[4,5-i]phenanthridine). These structures support the structures deduced by chemical and spectroscopic methods. The conformations of **3** and **4** are shown in *Fig. 2*. Since the *cis*-fused *B/C*-ring juncture of the hexahydrobenzo c phenanthridine alkaloid is flexible, two types of conformers are possible, *i.e.*, the *anti* and *syn* type [12]. By looking down to C(13) from C(14) in the Newman projection, C(17) of ring A adopts either a τ of ca. 180° or ca. 90° with respect to C(16) of ring D for an *anti*-periplanar and *syn*-periplanar conformation, respectively (Fig. 1). However, the crystal structures show both the same anti-cis B/Cring conformation with $\tau - 171.0^{\circ}$ for **3** and -174.8° for **4**. The ring-*B* conformations of 3 and 4 are half-chairs, and their C rings are almost planar due to the presence of two C=C bonds in this ring.

The dihedral angle between rings A and D in **3** is 62.3° . Although this value is slightly different from 46.79° in **1**, it is within the usual range of an alkaloid in the *anti*-type *B/C-cis*-ring conformation, and the relative positional relation of rings A and D in **3** is essentially the same as in **1**. The increased value of the dihedral angle of **3** by *ca*. 15° as compared with **1** could result from the lack of an intramolecular N(5) \cdots HO-C(12) H-bond, which is present in **1** [14].

The dihedral angle between rings A and D in **4** is 15.3° , pointing to a rather planar shape of this partial structure. This value is in the same range as that of 14-epicorynoline (10.5°) [15], which has a *trans-B/C*-ring junction being an 14-epimer of **1**. The torsion angle C(11)-C(13)-C(14)-C(16) is $4.9(3)^{\circ}$ for **4** and $-50.3(2)^{\circ}$ for **3**,

¹⁾ Trivial atom numbering; for systematic names of **3** and **4**, see *Exper. Part.*

	3	4
Crystallized from	CHCl ₃ /MeOH	acetone/MeOH
Empirical formula	$C_{21}H_{19}NO_4$	$C_{20}H_{17}NO_4$
$M_{\rm r}$ [g/mol]	349.37	335.35
Temperature [K]	120	120
Crystal system	monoclinic	orthorhombic
Space group	$P2_{1}/c$	$P2_{1}2_{1}2_{1}$
Z	4	4
θ Range for data collection [°]	1.99-28.26	1.87-28.23
Unit cell parameters:		
a [Å]	10.452(5)	6.2344(7)
b [Å]	12.839(5)	13.4636(16)
c [Å]	12.490(5)	18.588(2)
α [°]	90.000	90.000
β [°]	101.920(5)	90.000
γ [°]	90.000	90.000
V [Å ³]	1639.9(12)	1560.2(3)
$D_{\text{calc.}} \left[\text{g/cm}^{-3} \right]$	1.4151	1.428
$\mu(MoK_a) [mm^{-1}]$	0.098	0.100
F(000)	736	704
Radiation Mo $K_{\alpha} \lambda$ [Å]	0.71073	0.71073
No. of reflections collected	10476	10060
No. of independent reflections	3894	3690
Data, restraints, parameters	3894, 0, 235	3690, 0, 226
Goodness-of-fit on F^2	1.138	1.160
R indices $(I > 2\sigma(I))$	$R^1 = 0.0575, wR^2 = 0.1392$	$R^1 = 0.0517, wR^2 = 0.1249$
R indices (all data)	$R^1 = 0.0628, wR^2 = 0.1426$	$R^1 = 0.0538, wR^2 = 0.1262$
$\Delta_{\rm max}/\sigma$	0.000	0.000
$\Delta \rho$ (max; min) [e Å ⁻³]	0.397; -0.205	0.434; -0.258
Measurement	Bruker SMART APEX	Bruker SMART APEX
Program system	Bruker SMART	Bruker SMART
Structure determination	SIR92 [16]	SIR92 [16]
Refinement	full matrix SHELXL-97 [17]	full matrix SHELXL-97 [17]

Table 1. Crystallographic Data of 3 and 4

indicating a notable difference. This difference is in part due to the planar shape of ring C in 4 but not in 3. Also, the different direction of the Me-N(5) bond with respect to ring B would cause such a difference. The torsion angle C(22)-N(5)-C(14)-C(16) is $-47.9(2)^{\circ}$ in 3 due to the equatorial position of Me-N(5), similar to 1, and the corresponding C(21)-N(5)-C(14)-C(16) is $69.2(2)^{\circ}$ in 4 due to an axial Me-N bond, similar to 14-epicorynoline, endowed with a *trans-B/C*-ring juncture.

The conformations of **3** and **4** in solution were examined by NMR analyses (¹H at 499.7 and ¹³C at 125.7 MHz). The ¹H- and ¹³C-NMR data of **3** and **4** are given in *Tables 3* and 4, respectively. In the NOESY plot of **3**, the two NOEs Me-N(5)/H-C(4) and H-C(10)/Me-C(13) are observed (*Fig. 3,a*), suggesting that the corresponding H-atoms are located in close vicinity; the X-ray distance between these H-atoms amounts to 2.91 and 2.15 Å, respectively. The NOESY plot of **4** shows a NOE H-C(10)/H-C(11) (*Fig. 3,b*), the distance between these H-atoms being 2.22 Å in the

Bond lengths [Å]	3	4
C(11)-C(12)	1.329(3)	1.495(3)
C(11) - C(13)	1.521(2)	1.334(3)
C(13) - C(14)	1.547(2)	1.520(3)
C(13)-C(21)	1.546(2)	-
N(5)-C(6)	1.460(2)	1.460(2)
N(5)-C(14)	1.478(2)	1.483(2)
Bond angles [°]	3	4
C(12)-C(11)-C(13)	122.0(2)	124.8(2)
C(11) - C(13) - C(14)	108.9(1)	121.7(2)
C(11) - C(13) - C(17)	109.7(1)	124.2(2)
C(11) - C(13) - C(21)	109.3(1)	-
C(13) - C(14) - N(5)	110.0 (1)	111.5(2)
C(16) - C(14) - C(13)	110.0(1)	114.8(2)
C(16) - C(14) - N(5)	110.5(1)	110.3(2)
C(6) - N(5) - C(14)	109.7(1)	109.3(2)
C(6)-N(5)-C(22)	108.3(1)	-
C(6) - N(5) - C(21)	-	110.9(2)
C(14) - N(5) - C(22)	113.6(1)	-
C(14) - N(5) - C(21)	_	112.6(2)
Torsion angles [°]	3	4
C(12)-C(11)-C(13)-C(14)	31.0(2)	4.1(3)
C(11)-C(13)-C(14)-N(5)	71.6(2)	131.3(2)
C(11)-C(13)-C(14)-C(16)	-50.3(2)	4.9(3)
C(17)-C(13)-C(14)-N(5)	-49.0(2)	-48.4(2)
C(17) - C(13) - C(14) - C(16)	-171.0(1)	-174.8(2)
C(21) - C(13) - C(14) - C(16)	68.1(2)	-
C(21)-C(13)-C(14)-N(5)	-169.9(1)	-
C(21)-N(5)-C(14)-C(16)	-	69.2(2)
C(22) - N(5) - C(14) - C(16)	-47.9(2)	-

Table 2. Selected Bond Lengths, Bond Angles, and Torsion Angles of 3 and 4

crystal structure. That is to say, the conformations of **3** and **4** estimated from the ¹H-NMR spectra agree well with the X-ray crystal structures.

In the *B/C-cis*-type hexahydrobenzo[*c*]phenanthridine-type alkaloids, the equatorial position of the Me–N group with respect to ring *B* could be energetically stable because the steric hindrance with the neighboring atom in the case of axial position is avoided [14]. Therefore, when the Me–N group is in axial position, a downfield shift by $\Delta\delta$ ca. 0.5 could be expected for H–C(4), as compared with the case of Me–N in equatorial position, due to the effect of the lone electron pair of the N-atom on the electron shielding of C(4). The chemical shift of H–C(4) of **4** (δ 7.173) is shifted by $\Delta\delta$ 0.4 towards lower field, as compared with that of **3** (δ 6.80). This explains that the axial Me–N(5) in **4** results from the avoidance of the steric interaction between Me–N(5) and H–C(4).

The C(4) signal of **4** in the ¹³C-NMR spectrum is shifted to higher field ($\Delta\delta$ 4.3) compared to **3**. This is due to the effect on C(4) by the lone electron pair of N(5), when



Fig. 2. ORTEP Plots [13] of the molecular structures of a) 3^{1} and b) 4^{1} (50% probability ellipsoids)

Me-N(5) is in axial position. The chemical shift values of C(10) and C(11) of **4** are at higher field by $\Delta\delta$ 3.5 and 18.8, respectively, than those of **3**. This could also be due to a *gauche* effect. In conclusion, both ¹³C- and ¹H-NMR spectra show analogous tendencies concerning the conformational properties of **3** and **4**.

In summary, the present work established the most stable conformations of 3 and 4 in the crystal and solution states. The steric difference between the conformations of 3 and 4 could cause the different interaction with the adhesion protein ICAM-1, leading to the different inhibitory effects for cell adhesion.

	3	4
H-C(1)	6.62 (s)	6.60 (s)
H-C(4)	6.80 (s)	7.17 (s)
H-C(9)	6.79 (d, J = 8.0)	6.74 (d, J = 8.3)
H - C(10)	6.93 (d, J = 8.0)	7.16 (d, J = 8.0)
$CH_2(6)$	3.51 (d, J = 15.5), 3.99 (d, J = 15.5)	3.93 (d, J = 17.0), 4.45 (d, J = 17.0)
H - C(11)	5.83 (dd, J = 1.5, 9.5)	6.50(m)
$H-C(12)$ or $CH_2(12)$	6.27 (d, J = 9.5)	3.44 (<i>m</i>), 3.56 (<i>m</i>)
H - C(14)	3.19 (s)	4.56 (<i>m</i>)
Me-N(5)	2.12 (s)	2.00(s)
Me-C(13)	1.25 (s)	_
$CH_2(19), CH_2(20)$	5.95 (dd, J = 13, 1.5), 5.97 (s)	5.93 (dd, J = 9.5, 1.5), 5.98 (dd, J = 9.5, 1.5)

Table 3. ¹*H*-*NMR Data* (500 MHz, CDCl₃) of **3** and **4**¹). δ in ppm, *J* in Hz.

Table 4. ¹³C-NMR Data of **3** and **4** at 125 MHz in $CDCl_3^{1}$). δ in ppm, J in Hz.

	3	4
C(21)	25.066	_
Me-N(5)	42.144	34.565
C(1)	106.610	106.914
C(4)	112.341	108.028
C(9)	106.536	106.892
C(10)	119.520	116.054
C(6)	52.947	51.966
C(11)	138.175	119.383
C(12)	123.725	30.463
C(13)	39.563	126.639
C(14)	69.873	58.130
C(19)	101.024	100.738
C(20)	101.186	101.286
C(15)	127.224	127.350
C(16)	126.221	127.350
C(17)	135.054	128.542
C(18)	116.425	114.650
C(2)	147.453	146.350
C(3)	146.168	146.657
C(7)	142.984	144.502
C(8)	144.858	146.820

Experimental Part

 (\pm) -11,12-Didehydro-11-deoxycorynoline $(=(\pm)$ -(5bRS,12bRS)-5b,12b,13,14-Tetrahydro-5b,13-dimethyl[1,3]benzodioxolo[5,6-c]-1,3-dioxolo[4,5-i]phenanthridine; **3**) and (+)-11,13-Didehydro-11-deoxychelidonine (=(+)-rel-(12bR)-7,12b,13,14-Tetrahydro-13-methyl[1,3]benzodioxolo[5,6-c]-1,3-dioxolo[4,5-i]phenanthridine; **4**). (\pm) -Corynoline $((\pm)$ -**1**) was isolated from *Corydalis incise* (Papaveraceae) [3]. To a soln. of **1** (100 mg) in abs. benzene (4 ml), SOCl₂ (0.2 ml) was added. The mixture was refluxed for 30 min. The solvent was evaporated and H₂O (10 ml) was added to the residue. The aq. soln. was adjusted to pH 10 with 28% aq. NH₃ soln. and extracted with Et₂O (2 × 25 ml). The Et₂O layer was dried (Na₂SO₄) and concentrated to a yellow-orange oil, which was subjected to column chromatography



 $(Al_2O_3, benzene)$: **3** (64 mg). Prismatic crystals after recrystallization from CHCl₃/MeOH. M.p. 157.4–158.7°.

(+)-Chelidonine (2) was isolated from *Chelidonium majus* (Papaveraceae) [4]. Compound 4 was synthesized from 2 as described for 3: 4 as prismatic crystals after crystallization from acetone/MeOH [7]. M.p. $170.4-171.9^{\circ}$.

X-Ray Crystal Analyses of 3 and 4. A crystal was mounted on a nylon loop with 30% glycerol/mother liquor and then flash-frozen under a N₂ stream (120 K). Data collection was performed on a *CCD* diffractometer (*Bruker AXS SMART APEX*). The crystallographic data of 3 and 4 are given in *Table 1*. The crystal structure was solved by direct methods with the SIR92 program [16]. The positional parameters of the non-H-atoms were refined by a full-matrix least-squares method with anisotropic thermal parameters and the SHELXL-97 program [17], while those of the H-atoms were calculated on the basis of their configurational requirement. They were treated as riding with fixed isotropic displacement parameters ($U_{iso} = 1.2 U_{eq}$ for the associated C- or N-atoms, or $U_{iso} = 1.5 U_{eq}$ for O-atoms) and were not included as variables for the refinements. CCDC-724393 and 724392 contain the supplementary crystallographic data for 3 and 4. These data can be accessed free of charge *via* http://www.ccdc.cam.ac.uk/data_request/cif.

NMR Measurement. ¹H- and ¹³C-NMR Spectra: *Varian-VNMRS-500* (¹H at 499.7 MHz and ¹³C at 125.7 MHz) spectrometer; at 23° and at a concentration of 0.04M in CDCl₃ soln; D resonance of CDCl₃ as the lock signal; chemical shifts δ with respect to the internal reference Me₄Si; signal assignments by gCOSY, NOESY, gHMBC, gHSQC, and DEPT, where the estimated standard deviations are 0.001 ppm for the chemical shift and *ca.* 0.5 Hz for the coupling constant *J*. The nuclear *Overhauser* enhancement and exchange spectroscopy (NOESY) plot was recorded in the phase-sensitive mode at 23° and with a mixing time of 700 ms.

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